# Fluorine-19 Solid-State NMR Magic-Angle-Turning Experiments Using Multiple-Pulse Homonuclear Decoupling 

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For compounds giving "crowded" 1-dimensional magic-anglespinning spectra, information about the local atomic environment in the form of the chemical shift anisotropy (CSA) is sacrificed for high resolution of the less informative isotropic chemical shift. Magic-angle-turning (MAT) NMR pulse sequences preserve the C SA information by correlating it to the isotropic chemical shift in a 2 -dimensional experiment. For low natural abundance nuclei such as ${ }^{13} \mathrm{C}$ and ${ }^{15} \mathrm{~N}$ and under ${ }^{1} \mathrm{H}$ heteronuclear dipolar decoupling conditions, the dominant NMR interaction is the chemical shift. For abundant nuclei such as ${ }^{1} \mathrm{H},{ }^{19} \mathrm{~F}$, and ${ }^{31} \mathrm{P}$, the homonuclear dipolar interaction becomes a significant contribution to the observed linewidth in both $F_{1}$ and $F_{2}$ dimensions. We incorporate MREV8 homonuclear multiple-pulse decoupling sequences into the MAT experiment to give a multiple-pulse MAT (MPMAT) experiment in which the homonuclear dipolar interaction is suppressed while maintaining the chemical shift information. Extensive use of computer simulation using GAMMA has guided the pulse sequence development. In particular, we show how the MREV8 pulses can be incorporated into a quadrature-detected sequence such as MAT. The MP-MAT technique is demonstrated for a model two-site system containing a mixture of silver trifluoroacetate and calcium difluoride. The resolution in the isotropic evolution dimension is improved by faster sample spinning, shorter MREV8 cycle times in the evolution dimension, and modifications of the MAT component of the pulse sequence. © 1999 Academic Press

Key Words: fluorine-19; magic-angle turning; homonuclear decoupling; solid state; pulse sequence.

## INTRODUCTION

Valuable electronic, structural, and dynamic information about molecules in the solid state is obtained from the NMR chemical shift anisotropy (CSA). For powdered solids, the CSA is characterized experimentally by powder patterns for stationary samples or by the amplitude and frequency of the spinning sideband manifolds obtained under magic-angle-spinning (MAS) conditions. While CSA determinations are relatively straightforward for samples containing isolated equivalent nuclei, powder pattern and spinning sideband manifold overlap complicates similar CSA measurements for multiple-

[^0]site systems. To address this problem of spectral overlap, 2-dimensional NMR techniques have been developed to correlate the CSA to the isotropic chemical shift (1). Two strategies have been employed (2): the first correlates the isotropic chemical shift acquired under MAS conditions to the CSA collected in the $t_{\mathrm{b}}$ evolution dimension, while the second correlates the CSA $t_{\mathrm{a}}$ acquisition dimension to the isotropic chemical shift developed during the $t_{\mathrm{b}}$ evolution dimension. Into this latter category falls the magic-angle-turning (MAT) experiment.

In MAT and the related magic-angle-hopping (MAH) (3) experiments, the sample is moved in three, equally spaced successive steps about the magic angle so that static (MAH) or pseudo-static CSA (MAT) spectra are collected in the $t_{\mathrm{a}}$ acquisition dimension (Fig. 1). Each evolution increment $\Delta t_{\mathrm{b}}$ is divided into three time segments $\Delta t_{\mathrm{b}} / 3$, separated in sample orientation by $120^{\circ}$, which averages the principal components of the chemical shift to the isotropic value. The difference between the two experiments lies in how this sample movement is effected: the MAH variant employs discrete "hops" whereas the MAT experiment employs very slow continuous turning ( $\nu_{\text {rot }}=30-600 \mathrm{~Hz}$ ) of the sample. For the MAT experiment, the start of each evolution increment $\Delta t_{\mathrm{b}} / 3$ is rotor-synchronized and begins at times $0,1 / 3 \tau_{\text {rot }}$, and $2 / 3 \tau_{\text {rot }}$ where $\tau_{\text {rot }}=1 / \nu_{\text {rot }}=$ the rotor period.

MAT experiments incorporating high-power ${ }^{1} \mathrm{H}$ decoupling and ${ }^{1} \mathrm{H}$ cross-polarization techniques have been applied successfully to dilute spin- $1 / 2$ nuclei such as ${ }^{13} \mathrm{C}$ and ${ }^{15} \mathrm{~N}(4-12)$. CSA values in solid organic samples containing up to 15 different ${ }^{13} \mathrm{C}$ nuclei have been resolved (10). However, current MAT techniques cannot be applied directly to samples containing abundant spin- $1 / 2$ nuclei such as ${ }^{1} \mathrm{H},{ }^{19} \mathrm{~F}$, and ${ }^{31} \mathrm{P}$ since the large homonuclear dipolar interaction present in such systems obscures the chemical shift information. The inability to measure the chemical shift in the presence of such homonuclear dipolar interactions hinders the determination of ${ }^{1} \mathrm{H}$, ${ }^{19} \mathrm{~F}$, and ${ }^{31} \mathrm{P}$ CSA values in systems for which there may already be limited structural, electronic, and dynamic information. Materials for which diffraction techniques in particular are limited and which would thus benefit from CSA characterization include fluorine- and proton-containing polymers $(2,13)$, inorganic glasses (14), mesoporous materials (15), and
phosphorus-containing bioorganic solids (16). Furthermore, we note that the electronic features of hydrogen-bonded systems are accessible from the ${ }^{1} \mathrm{H}$ CSA values $(17,18)$.

In this paper, we present the incorporation of multiple-pulse (MP) homonuclear dipolar decoupling into MAT NMR sequences to effect the removal of the homonuclear dipolar interaction in order to obtain high-resolution chemical shift and CSA correlation spectra. The strategy of combining the MREV8 multiple-pulse sequence into a MAT experiment is developed and then demonstrated for one- and two-site ${ }^{19} \mathrm{~F}$ containing samples. We show that a direct combination of the MREV8 and MAT sequences is not possible, but can be successfully implemented with appropriate modification of the MREV8 sequence. Finally, strategies to enhance the resolution in the isotropic evolution $t_{b}$-dimension are presented and evaluated.

## EXPERIMENTAL

Microcrystalline solids of silver trifluoroacetate $\left(\mathrm{AgO}_{2} \mathrm{CCF}_{3}\right.$, Aldrich) and calcium difluoride ( $\mathrm{CaF}_{2}$, Aldrich) were used without any additional purification.

The NMR spectra were collected on a Chemagnetics CMX200 spectrometer operating at 200.13 MHz for protons and 188.29 MHz for fluorine. A commercially available Chemagnetics H-F double-resonance APEX spinning probe was used with $4-\mathrm{mm}$ (od) pencil zirconia rotors, together with fluorine-free Vespel drive tips and spacer inserts. In order to minimize both the $\mathbf{B}_{0}$ and $\mathbf{B}_{1}$ field inhomogeneities the sample was confined to the central 2 mm of the rotor. The probe magic angle was set by minimizing the ${ }^{19} \mathrm{~F}$ linewidth of $p$-tert-butylcalix[4]arene- $\alpha, \alpha, \alpha$-trifluorotoluene (19).

For the turning experiments requiring low spinning rates ( $\nu_{\text {rot }}$ $=30-600 \mathrm{~Hz}$ ), a special drive tip without flutes was used in conjunction with a special brass constrictor tube $(0.8 \mathrm{~mm}$ (id), 26 mm long) placed in the line carrying the drive air. Sample spinning was controlled by manual adjustment of the drive and bearing air pressures for frequencies $\nu_{\text {rot }}<300 \mathrm{~Hz}$ and automatically for spinning frequencies $>300 \mathrm{~Hz}$. Air pressures for the bearing and drive of 10 and 2 psi , respectively, give a spinning rate of $\sim 50 \mathrm{~Hz}$ which remains constant ( $\pm 1 \mathrm{~Hz}$ ) over the course of the experiments (up to 12 h ). In contrast to other MAT-type experiments, the sample spinning is not placed under an electronic feedback control system (5). A Bruker CXP amplifier operating in class C mode was used for ${ }^{19} \mathrm{~F}$ RF irradiation. All experiments were carried out at ambient temperatures.

The standard multiple pulse probe tune-up for ${ }^{19} \mathrm{~F}$ was followed (20). The 45, 90, 135, 180, 225, 270, and $310^{\circ}$ pulses were set using a sample of $\mathrm{C}_{6} \mathrm{~F}_{6}$ held in a glass sphere and placed at the center of the probe coil. The 1-D MREV8 spectra were collected using the quadrature-detected pulse sequence of Barbara and Baltusis (21). MREV8 acquisition cycle times of $t_{\mathrm{c}}=36$ to $40.8 \mu \mathrm{~s}$ were used (corresponding to $\tau$ values of 3.0 to $3.4 \mu \mathrm{~s}$ ), with $90^{\circ}$ pulse durations of 1.36 to $1.45 \mu \mathrm{~s}$. An
integral multiple of the MREV8 acquisition cycle was used for the evolution dwell time in order to minimize the spectral width in the $t_{1}$ dimension. The scaling of the chemical shift in the evolution dimension was calibrated using a sample of silver trifluoroacetate (AgTFA) by collection of data with the transmitter offset at $1-\mathrm{kHz}$ increments from the isotropic peak at 188.2965514 MHz . Further NMR experimental details are given in the appropriate figure captions. $T_{1}$ measurements were carried out under MREV8 conditions using the satura-tion-recovery method: AgTFA (1.2 s) and calcium difluoride ( $>20 \mathrm{~s}$ ).

The computer simulations were carried out using GAMMA version $3.5 \beta$ (22) on a Sun ultrasparc1 workstation. Simulation of a two-site, 2-D MP-MAT spectrum took approximately 12 h . The simulated data were output in the Chemagnetics Spinsight data format and are processed in the same way as experimental data.

## RESULTS AND DISCUSSION

## (a) The Magic-Angle-Hopping and -Turning Experiments

The theory for MAH and MAT has been described in detail in previous publications $(1,3,5)$. For now, we will consider a brief overview of the theory and features of both experiments relevant to the incorporation of multiple-pulse sequences. In particular, the MAH sequence is examined and developed in the context of the GAMMA simulation work. The relevant features of the MAH sequence are then brought to bear on the subsequent development of the MAT experiments.

The generalized MAT pulse sequence is depicted in Fig. 1. The pulses are synchronized to the rotor orientation as it rotates about the magic angle. The first segment starts at time $\tau_{\text {rot }}=0$, at which point the magnetization evolves for time $t_{\mathrm{b}} / 3$ through an angle $\Phi_{1}$ in the $x y$ plane. Selection of either the cosine or the sine component by a projection pulse $p_{1}$ stores the magnetization along the longitudinal axis for the duration, $L$, of this time segment. The remaining transverse component is purged by $T_{2}$ relaxation and dispersion (note that $T_{1} \geqslant T_{2}$ ). After a time of $\tau_{\text {rot }} / 3$, the sample has rotated by $120^{\circ}$, and a tipping pulse $r_{2}$ allows the magnetization to evolve in the $x y$ plane for time $t_{\mathrm{b}} / 3$ through an angle $\Phi_{2}$; one of the transverse components is selected $\left(p_{2}\right)$ in a similar manner. This is repeated for the final time period, at which point the FID is acquired. Since the magnetization has evolved under the chemical shift interaction in three mutually perpendicular directions (when viewed from the sample), the summed signal gives the isotropic chemical shift according to

$$
\begin{equation*}
\omega_{x}+\omega_{y}+\omega_{z}=3 \omega_{\mathrm{i} \mathrm{so}} . \tag{1}
\end{equation*}
$$

In a collection of a 2 -dimensional hypercomplex data set, Eq. [1] holds true for specific values of the projection pulse phases $p_{\mathrm{i}}$ and appropriate combinations of the respective FIDs. A


FIG. 1. Pulse sequence for the 2-dimensional magic-angle-turning (MAT) NMR experiment: (a) basic MAT pulse sequence (1) which becomes the multiple-pulse MAT (MP-MAT) sequence upon addition of MREV8 homonuclear decoupling during the (b) evolution and (c) acquisition dimensions. All pulses are $90^{\circ}$ except for $c\left(45^{\circ}\right)$ and $r_{\phi}$ (see text); L refers to the magnetization storage period, $r_{\mathrm{i}}$ are the read pulses, and $p_{\mathrm{i}}$ are the projection pulses. In (b) the number of MREV8 cycles per evolution dwell time increment, $\Delta \mathrm{t}_{\mathrm{a}}$, is m , while n is the number of evolution increments. In (c) the arrow indicates the sampling window and al is the number of collected acquisition points.

2-dimensional spectrum with an isotropic chemical shift dimension is produced from a set of FIDs in which the amplitude is modulated by the functions $\cos \left(\Phi_{1}+\Phi_{2}+\Phi_{3}\right)$ and $\sin \left(\Phi_{1}\right.$ $\left.+\Phi_{2}+\Phi_{3}\right)$. Eight combinations of the phases of the three projection pulses $p_{\mathrm{i}}$ :

$$
\begin{align*}
& F_{1}\left(t_{\mathrm{b}}, t_{\mathrm{a}}\right)=\cos \left(\Phi_{1}\right) \times \cos \left(\Phi_{2}\right) \times \cos \left(\Phi_{3}\right) F_{\mathrm{a}}\left(t_{\mathrm{a}}\right)  \tag{2}\\
& F_{2}\left(t_{\mathrm{b}}, t_{\mathrm{a}}\right)=-\sin \left(\Phi_{1}\right) \times \sin \left(\Phi_{2}\right) \times \cos \left(\Phi_{3}\right) F_{\mathrm{a}}\left(t_{\mathrm{a}}\right)  \tag{3}\\
& F_{3}\left(t_{\mathrm{b}}, t_{\mathrm{a}}\right)=-\sin \left(\Phi_{1}\right) \times \cos \left(\Phi_{2}\right) \times \sin \left(\Phi_{3}\right) F_{\mathrm{a}}\left(t_{\mathrm{a}}\right)  \tag{4}\\
& F_{4}\left(t_{\mathrm{b}}, t_{\mathrm{a}}\right)=-\cos \left(\Phi_{1}\right) \times \sin \left(\Phi_{2}\right) \times \sin \left(\Phi_{3}\right) F_{\mathrm{a}}\left(t_{\mathrm{a}}\right)  \tag{5}\\
& F_{5}\left(t_{\mathrm{b}}, t_{\mathrm{a}}\right)=\sin \left(\Phi_{1}\right) \times \cos \left(\Phi_{2}\right) \times \cos \left(\Phi_{3}\right) F_{\mathrm{a}}\left(t_{\mathrm{a}}\right)  \tag{6}\\
& F_{6}\left(t_{\mathrm{b}}, t_{\mathrm{a}}\right)=\cos \left(\Phi_{1}\right) \times \sin \left(\Phi_{2}\right) \times \cos \left(\Phi_{3}\right) F_{\mathrm{a}}\left(t_{\mathrm{a}}\right)  \tag{7}\\
& F_{7}\left(t_{\mathrm{b}}, t_{\mathrm{a}}\right)=\cos \left(\Phi_{1}\right) \times \cos \left(\Phi_{2}\right) \times \sin \left(\Phi_{3}\right) F_{\mathrm{a}}\left(t_{\mathrm{a}}\right)  \tag{8}\\
& F_{8}\left(t_{\mathrm{b}}, t_{\mathrm{a}}\right)=-\sin \left(\Phi_{1}\right) \times \sin \left(\Phi_{2}\right) \times \sin \left(\Phi_{3}\right) F_{\mathrm{a}}\left(t_{\mathrm{a}}\right) \tag{9}
\end{align*}
$$

generate the required cosine- and sine-modulated FIDs:

$$
\begin{align*}
& F_{1}+F_{2}+F_{3}+F_{4}=\cos \left(\Phi_{1}+\Phi_{2}+\Phi_{3}\right) F_{\mathrm{a}}\left(t_{\mathrm{a}}\right)=F_{\text {real }} \\
& F_{5}+F_{6}+F_{7}+F_{8}=\sin \left(\Phi_{1}+\Phi_{2}+\Phi_{3}\right) F_{\mathrm{a}}\left(t_{\mathrm{a}}\right)=F_{\text {imag }} \tag{10}
\end{align*}
$$

where $F_{\mathrm{a}}\left(t_{\mathrm{a}}\right)$ is the acquired complex FID.
Table 1 collects the basic pulse phases for the prototype full MAH or MAT sequence (after Hu et al. (1)). Only the phases of the first projection pulse $p_{1}$ and the receiver differ between the two $F_{\text {real }}$ and $F_{\text {imag }}$ data sets. In a hypercomplex MAT experiment $F_{\text {real }}$ and $F_{\text {imag }}$ are collected sequentially and combined in subsequent processing to give an absorption-absorp-tion-phased 2-dimensional spectrum.

The full MAT pulse sequence (Fig. 1a) has two disadvantages for spin-dilute nuclei ( 1,5 ): First, loss of magnetization due to three longitudinal storage periods lowers the signal-tonoise ratio. Second, acquisition begins immediately after the final read pulse $r_{\phi}$ and may be adversely influenced by probe dead-time and receiver ring-down effects. The incorporation of the third evolution period into the acquisition period both improves the signal-to-noise ratio and reduces baseplane distortions (4). While the signal-to-noise ratio of the full MAT sequence may be a disadvantage for low natural abundance nuclei, it is less of a disadvantage for the higher sensitivity and natural abundance of ${ }^{1} \mathrm{H},{ }^{19} \mathrm{~F}$, and ${ }^{31} \mathrm{P}$. Furthermore, baseplane distortions are reduced since external RF filters are not required and the probes are designed to have short ring-down. We begin by incorporating a MREV8 multiple-pulse sequence into the full MAH and MAT pulse sequences described in Fig. 1 and conclude with some of the pulse sequence improvements mentioned above.

## (b) GAMMA Simulation of the MAH Experiment

Theoretical considerations. The introduction of a multi-ple-pulse homonuclear decoupling sequence within the MAT

TABLE 1
The Basic Phase Table for the MAH and MAT Pulse Sequences in Fig. 1

| $n t$ | $\begin{aligned} F_{\text {real }}= & F_{1}+F_{2}+F_{3} \\ & +F_{4} \end{aligned}$ |  |  |  | $\begin{aligned} F_{\text {imag }}= & F_{5}+F_{6}+F_{7} \\ & +F_{8} \end{aligned}$ |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 |
| $r$ | $Y$ | $Y$ | Y | Y | Y | $Y$ | Y | $Y$ |
| $p_{1}$ | $X$ | $Y$ | Y | $X$ | Y | $X$ | X | $Y$ |
| $p_{2}$ | X | Y | X | $Y$ | X | $Y$ | X | $Y$ |
| $p_{3}$ | $X$ | X | Y | Y | $X$ | $X$ | Y | Y |
| $r c \nu r$ | $X$ | $\bar{X}$ | $\bar{X}$ | $\bar{X}$ | $\bar{X}$ | $\bar{X}$ | $\bar{X}$ | $X$ |
| FID | $F_{1}$ | $F_{2}$ | $F_{3}$ | $F_{4}$ | $F_{5}$ | $F_{6}$ | $F_{7}$ | $F_{8}$ |

[^1]experiment has been achieved through the extensive use of computer simulations. The object-oriented GAMMA (22) package was used to simulate various aspects of the experiment using a density matrix approach. In the MAT experiment, the rotor in which the sample is packed is slowly rotated about the magic angle in order to spatially select the three components of the chemical shift tensor during the $t_{\mathrm{b}}$ evolution periods. Therefore, the Hamiltonian describing the system is time dependent, continuously changing as the rotor turns. To simulate such an experiment using a density matrix approach requires that the rotor period be split up into small increments, over which the Hamiltonian can be assumed constant. For each increment the Hamiltonian is recalculated and the density matrix evolved. Alternatively, a solution based on Floquet methods $(23,24)$ could be pursued to render the Hamiltonian constant over the full rotor period. However, it was felt that both methods would be too complicated and computer intensive to implement successfully. Therefore, instead of attempting to simulate the turning experiment of Gan (4), it was decided to investigate the possibilities of simulating the original hopping (MAH) experiment of Bax and co-workers (3).

The MAH experiment is essentially a static experiment, and therefore the Hamiltonian describing the system is time independent. Evolution under the three different components of the chemical shift tensor is brought about by a rapid rotation of the sample by $120^{\circ}$ about the magic angle at the end of the evolution period. In a simulation, therefore, a constant Hamiltonian can be used to propagate the density matrix during each third of the MAH experiment. This reduces the complexity of the simulation and also decreases the execution time. The main purpose of attempting to simulate the MAH experiment is to guide the incorporation of a multiple-pulse sequence into the evolution and detection periods of the MAT experiment. At this stage the effectiveness of the multiple-pulse sequence to remove the homonuclear dipolar interaction is not under investigation, and therefore a system consisting of isolated equivalent spins was used. The system then evolves under a chemical shift Hamiltonian alone. However, we have used two mutually isolated spin baths, each consisting of an ensemble of equivalent ${ }^{19} \mathrm{~F}$ nuclei (ignoring both dipolar and indirect coupling) as the basis of the 2-D simulations calculated in this paper to show more clearly how the phase cycling of the final pulse sequence behaves. This case will be referred to as a two-spin system.

In the computer simulation, the total Hamiltonian, $H_{\text {Tот }}$ for the two-spin system is

$$
\begin{equation*}
H_{\text {тот }}=H_{Z}+H_{\mathrm{cs}}^{1}+H_{\mathrm{cs}}^{2}, \tag{12}
\end{equation*}
$$

where $H_{\mathrm{Z}}$ is the Zeeman Hamiltonian and $H_{\mathrm{cs}}^{1}$ and $H_{\mathrm{cs}}^{2}$ are the chemical shift Hamiltonians for the two spin types. In order to arrive at the final expressions for the Hamiltonian used in the simulation (which incorporates the spatial dependence of the motion of the sample about the magic angle), we have used
standard spherical tensor methods. We follow closely the work of Haeberlen (25) and Maricq and Waugh (26).

The individual terms of the Hamiltonian can be expressed as the product of second-rank tensors by the equation

$$
\begin{equation*}
H^{\lambda}=C^{\lambda} \sum_{l} \sum_{l=-m}^{l}(-1)^{m} R_{l,-m}^{\lambda} T_{l, m}^{\lambda}, \tag{13}
\end{equation*}
$$

where $C^{\lambda}$ comprises physical constants appropriate to the type of interaction $\lambda, R_{l,-m}$ is the spatial spherical tensor, and $T_{l, m}$ is the spin spherical tensor. The Zeeman Hamiltonian may be removed from the equation by transforming the spin tensor $T$ into a coordinate system rotating at the Larmor frequency about the $z$ axis in the laboratory frame. The periodic time dependence may be removed by average Hamiltonian theory (27), and to lowest order the Hamiltonian for a chemical shift interaction is

$$
\begin{equation*}
H_{0}^{\mathrm{CS}}=C^{\mathrm{CS}} \sum_{i=1}^{2}\left(R_{00}^{\mathrm{CS}} T_{00}^{\mathrm{CS}}+R_{20}^{\mathrm{CS}} T_{20}^{\mathrm{CS}}\right)_{i}, \tag{14}
\end{equation*}
$$

where $i$ is nuclear spin index. Both the spin and the spatial tensors are expressed in the laboratory frame. The spatial tensor is diagonal in its principal axis system (PAS) and can be related back to the laboratory frame by applying two successive rotational transformations so that the tensor goes through an intermediate reference frame fixed in the sample rotor. The $R_{00}$ component represents the isotropic chemical shift and is invariant under rotation. The $R_{20}$ components may be rotated using standard Wigner rotation matrices in the equation

$$
\begin{equation*}
R_{l, m}^{\lambda, i}=\sum_{m^{\prime}} D_{m^{\prime}, m}^{l}\left(\alpha^{\lambda}, \beta^{\lambda}, \gamma^{\lambda}\right) \rho_{l, m^{\prime}}^{\lambda, i}, \tag{15}
\end{equation*}
$$

where $l$ is the rank of the tensor, $\alpha, \beta$, and $\gamma$ are the Euler angles, and $\rho_{l, m}$ the components of the irreducible spherical tensor in the new reference frame. The definition of the Euler angle matrix elements is given by Spiess (28). With two rotations, for $R_{20}$ this is then

$$
\begin{equation*}
R_{2,0}^{\mathrm{CS}, i}=\sum_{m^{\prime}=-2}^{2} D_{m^{\prime}, 0}^{2}\left(0, \theta, \omega_{\mathrm{r}} t\right) \sum_{m^{\prime \prime}=-2}^{2} D_{m^{\prime}, m^{\prime}}^{2}(\alpha, \beta, \gamma) \rho_{2, m^{\prime \prime}}^{\mathrm{CS},} \tag{16}
\end{equation*}
$$

where the $\rho_{2 \pm 1}$ are zero for the chemical shift interaction. The angles $\alpha, \beta$ and $\gamma$ relate the principal axis system of the chemical shift tensor to the axis system fixed in the rotor spinner. The angles $\theta$ and $\omega_{\mathrm{r}} t$ relate the rotor-fixed axis system to the laboratory axis system. In the simulations, we restrict ourselves to an axially symmetric tensor, which allows the $\rho_{2 \pm 2}$
terms to be set to zero to reduce computational time. Thus, the only terms required in the simulations arise from $\rho_{20}$. Substitution of the correct Wigner matrix elements gives

$$
\begin{align*}
R_{2,0}^{\mathrm{CS}, i}= & \left(\frac{3 \cos ^{2} \theta-1}{2}\right)\left(\frac{3 \cos ^{2} \beta-1}{2}\right) \rho_{2,0}^{\mathrm{CS}, i} \\
& +\frac{3}{4} \sin ^{2} \theta \sin ^{2} \beta \cos \left(2 \omega_{\mathrm{r}} t+2 \gamma\right) \rho_{2,0}^{\mathrm{CS}, i} \\
& -\frac{3}{4} \sin 2 \theta \sin 2 \beta \cos \left(\omega_{\mathrm{r}} t+\gamma\right) \rho_{2,0}^{\mathrm{CS}, i} . \tag{17}
\end{align*}
$$

In the principal axis system of the chemical shift tensor $\rho_{2,0}^{\mathrm{CS}}=$ $\sqrt{3 / 2 \zeta}$ and $\rho_{0,0}^{\mathrm{CS}}=\delta_{\text {iso }}$, where $\delta_{\text {iso }}$ is the isotropic chemical shift and $3 \zeta / 2=\delta_{33}-\frac{1}{2}\left(\delta_{11}+\delta_{22}\right)$ the chemical shift anisotropy. For the spin tensor $T_{00}^{\mathrm{CS}, i}=I_{0}^{i} B_{0}$ and $T_{00}^{\mathrm{CS}, i}=\sqrt{2 / 3} I_{0}^{i} B_{0}$. On substitution into Eq. [14] one gets, in angular frequency units

$$
\begin{align*}
H_{0}=\sum_{i} & \left(\gamma_{i} \delta_{\text {iso }}^{i} I_{0} B_{0}+\gamma_{\mathrm{i}} \xi^{i}\left[\left(\frac{3 \cos ^{2} \theta-1}{2}\right)\left(\frac{3 \cos ^{2} \beta-1}{2}\right)\right.\right. \\
& +\frac{3}{4} \sin ^{2} \theta \sin ^{2} \beta \cos \left(2 \omega_{\mathrm{r}} t+2 \gamma\right) \\
& \left.\left.-\frac{3}{4} \sin 2 \theta \sin 2 \beta \cos \left(\omega_{\mathrm{r}} t+\gamma\right)\right] I_{0}^{i} B_{0}\right) . \tag{18}
\end{align*}
$$

When $\theta=54.7^{\circ}$ (the magic angle) and with $\omega_{0}=\gamma_{0} B_{0}$, Eq. [18] simplifies to

$$
\begin{gather*}
H_{0}=\sum_{i} \omega_{0}^{i}\left(\delta_{\text {iso }}^{i}+\zeta^{i}\left[\frac{3}{4} \sin ^{2} \theta \sin ^{2} \beta \cos \left(2 \omega_{\mathrm{r}} t+2 \gamma\right)\right.\right. \\
\left.\left.-\frac{3}{4} \sin 2 \theta \sin 2 \beta \cos \left(\omega_{\mathrm{r}} t+\gamma\right)\right]\right) I_{0}^{i} . \tag{19}
\end{gather*}
$$

Since the MAH experiments evolve under a time-independent Hamiltonian during the acquisition period, Eq. [19] can be simplified further, so that it is dependent only on one angle. This reduction is achieved by taking the PAS frame directly into the laboratory frame and bypassing the axis system fixed in the rotor. Equation [19] then becomes

$$
\begin{equation*}
H_{0}=\sum_{i} \omega_{0}^{i}\left(\delta_{\mathrm{iso}}^{i}+\zeta^{i}\left(\frac{3 \cos ^{2} \beta-1}{2}\right)\right) I_{0}^{i} . \tag{20}
\end{equation*}
$$

This simplification reduces the number of points required for an adequate powder average considerably. In the $t_{\mathrm{b}}$ dimension the powder average is performed over the two angles $\gamma$ and $\beta$. The crystallite orientations in the powder average were calculated using Hammersley points $(29,30)$. This method is comparable to those of Conroy (31) and Wolfsberg (32) but has the advantage that it is not restricted to specific set sizes.

MAH with MREV8 decoupling. It has been shown that the homonuclear dipolar couplings can be removed successfully by applying multiple-pulse decoupling techniques (33). Therefore, in order to use the MAT technique for abundant spins, an


FIG. 2. ${ }^{19} \mathrm{~F}$ MREV8 NMR spectrum ( 188.2976 MHz ) of a stationary sample of AgTFA illustrating $0^{\circ}$ (top) and $180^{\circ}$ (middle) receiver phase selection. The bottom trace is the summed residual. (a) Simulation using an axial CSA tensor (see text). (b) Experimental: 64 acquisitions; 6-s recycle delay; 256 points collected, and zero-filled to 512 ; pulses: $45^{\circ}=0.81 \mu \mathrm{~s}, 90^{\circ}$ $=1.36 \mu \mathrm{~s}, 135^{\circ}=1.91 \mu \mathrm{~s}, 225^{\circ}=2.99 \mu \mathrm{~s}, 270^{\circ}=3.53 \mu \mathrm{~s}, 315^{\circ}=4.26 \mu \mathrm{~s}$.
appropriate form of multiple-pulse selective averaging during the evolution and acquisition periods of the 2-D sequence must be introduced. We chose, in the first instance, to work with the MREV8 $(34,35)$ multiple-pulse sequence.

The introduction of the MREV8 pulse sequence into both the evolution and the detection periods of the MAT experiment raises two main problems. The first involves the correct phase cycling of the receiver during the detection period in order for the MAT sequence to work properly. The second problem arises during the evolution period and is one of retaining quadrature information when the different sine and cosine components are selected after the evolution period. Under the majority of multiple-pulse decoupling sequences the effective magnetization no longer precesses about the $z$ axis aligned along the $\mathbf{B}_{0}$ field, and therefore the sine and cosine components of the magnetization detected by the receiver in quadrature are no longer of the correct magnitude with respect to each other. If this is not corrected, quadrature ghosts will appear in the final 2-D spectrum.

In a MAT experiment two hypercomplex data sets are collected which are subsequently combined to form a single 2-dimensional spectrum. Each experiment, in its most simple form, consists of a four-pulse phase cycle (Table 1). For one
${ }^{19}$ F MAT EXPERIMENTS WITH MULTIPLE-PULSE DECOUPLING

TABLE 2
The Basic Phase Table for the MP-MAT Pulse Sequence in Fig. 1

| $n t$ | $F_{1}$ |  |  |  | $F_{2}$ |  |  |  | $F_{3}$ |  |  |  | $F_{4}$ |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 |
| $r$ | Y | Y | Y | $Y$ | Y | Y | Y | Y | Y | Y | Y | Y | Y | Y | Y | Y |
| c | $X$ | $X$ | $X$ | $X$ | X | X | X | X | X | X | X | X | $X$ | X | $X$ | X |
| $p_{1}$ | $X$ | $X$ | $X$ | $X$ | Y | Y | Y | Y | $Y$ | $Y$ | Y | Y | $X$ | X | $X$ | $X$ |
| $p_{2}$ | $X$ | $X$ | $X$ | $X$ | Y | Y | Y | Y | X | X | X | X | Y | Y | $Y$ | Y |
| $p_{3}$ | $X$ | $X$ | $X$ | $X$ | X | X | $X$ | $X$ | $X$ | $X$ | X | X | X | X | $X$ | $X$ |
| $r_{\text {¢ }}$ | $\{135$ | 45 | 90 | 90 | 315 | 225 | 270 | 270 | 315 | 225 | 270 | 270 | 315 | 225 | 270 | 270 |
|  | Y | $\bar{Y}$ | X | $\bar{X}$ | Y | $\bar{Y}$ | X | $\bar{X}$ | Y | $\bar{Y}$ | X | $\bar{X}$ | Y | $\bar{Y}$ | X | $\bar{X}$ |
| $r c \nu r$ | Y | $\bar{Y}$ | $X$ | $\bar{X}$ | Y | $\bar{Y}$ | X | $\bar{X}$ | Y | $\bar{Y}$ | X | $\bar{X}$ | Y | $\bar{Y}$ | $X$ | $\bar{X}$ |
|  | $F_{5}$ |  |  |  | $F_{6}$ |  |  |  | $F_{7}$ |  |  |  | $F_{8}$ |  |  |  |
| $n t$ | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 |
|  | Y |  |  |  |  |  |  |  |  | X | $X$ | X | Y | $Y$ | Y | Y |
| $r_{\phi}$ | 315 | 225 | 270 | 270 | 315 | 225 | 270 | 270 | 315 | 225 | 270 | 270 | 135 | 45 | 90 | 90 |

Note. The real (top) and imaginary (bottom) data sets are collected with variation of the phase of the projection pulse $p_{1}$ and the pulse durations (in degrees) of the final read pulse $r_{\phi} ; n t$ represents the transition number.
experiment, during the first acquisition the receiver is set to $0^{\circ}$ $(X)$, and for the next three acquisitions it is set to $180^{\circ}(\bar{X})$. In the second experiment, the first three acquisitions have the receiver set to $180^{\circ}$ and in the final acquisition the receiver is set to $0^{\circ}$. With the addition of the MREV8 sequence during the acquisition stage, one can no longer simply change the receiver phase to achieve the correct $180^{\circ}$ phase cycling since the precession of the magnetization is no longer along $\mathbf{B}_{0}$. In order to achieve the required phase cycling during acquisition, the final MAT read pulse $r_{\phi}$ must be replaced with a MREV8 sequence that preserves quadrature and can produce the necessary phase inversion of the signal.

Recently, various groups have introduced multiple-pulse sequences that preserve quadrature ( $21,36,37$ ). The MREV8 sequence introduced by Barbara and Baltusis (21) achieves a quadrature-detected spectrum by altering the magnitude and phase of the preparation pulses so that the magnetization precesses about different axes. When the resulting FIDs are coadded, they produce a FID which has equal magnitude signals in both real and imaginary channels. The published preparation pulse durations are $45_{270}^{\circ}, 135_{90}^{\circ}, 90_{0}^{\circ}$, and $90_{180}^{\circ}$, where the subscript denotes both the pulse and receiver phase. We show that if each preparation pulse duration is increased by $180^{\circ}$ so that the preparation pulses are now $225_{270}^{\circ}, 315_{90}^{\circ}, 270_{0}^{\circ}$, and $270_{180}^{\circ}$, then an inverted spectrum is achieved on Fourier transformation. Figure 2a shows two 1-dimensional simulated MREV8 spectra, where the lower spectrum has been inverted with respect to the top spectrum by applying the second set of preparation pulses. Figure 2 b shows the results of an actual implementation of the pulse sequence using a sample of AgTFA. The summation spectrum is given at the bottom of the figure, and it is clear that the relative phases of the two spectra
are very close to $180^{\circ}$. The inverted spectrum of the AgTFA shows a small zero-frequency spike at the transmitter, which may arise from the lower precision of the longer pulses used in the second MREV8 sequence.

More complex MAT sequences such as the MAT $\pm$ (6) sequence (discussed later), require receiver phase cycling in steps of $90^{\circ}$. We have been unable to find preparation pulse duration and phase combinations that would allow the MREV8 multiple-pulse sequence to mimic such receiver phase shifts. Therefore we have followed an alternate approach to meet the receiver phase inversion requirements of the initial MAT sequence which can also be used in more complex MAT experiments. The technique involves collecting and storing separately each individual FID that makes up the phase cycle. Each FID is collected with the same receiver phase. At the end of the experiment, the appropriate phase transformation is applied to the FIDs and then they are combined to give the real and imaginary data sets. This procedure replaces what a quadrature receiver in spectrometers does automatically during data acquisition. While the above approach generates a number of temporary files, it does allow the consistent use of the shorter preparation pulses, which in turn eliminates any residual summation signal (Fig. 2) due to the lower precision in the longer preparation pulses. The final result is a better addition of the MAT acquisition FIDs and an improved 2-D spectrum.

In each of the three evolution periods of the full MAT experiment, the MREV8 sequence is required to remove the homonuclear decoupling. At the start of each period the magnetization is rotated into the $x y$ plane by a single $90^{\circ}$ read pulse $r$ along the $y$ axis (see Fig. 1 and Table 2). At the end of each evolution period the sine or cosine projection of the magnetization is selected and stored along the [001] $z$ axis by applying
a $90^{\circ}$ pulse $p_{i}$ of the correct phase. This sequence again requires that the magnetization precess about the $z$ axis so that the magnitude of each projection is correct. However, the MREV8 sequence upsets this situation, since magnetization precesses about the [101] rather than the [001] axis, as discussed above. In order to retain the correct quadrature relationship, a compensation pulse $c$ is applied at the end of the MREV8 evolution period to rotate the magnetization precession back into the $x y$ plane (38). For the traditional MREV8 sequence, Jackson has shown that this can be achieved with a $45^{\circ}$ pulse applied about the $y$ axis (39). With the phase cycle of the MAT sequence, each preparation pulse is along the $y$ axis, that is $90^{\circ}$ out of phase from the normal MREV8 preparation pulse (21). Therefore, all the pulses used in the MREV8 sequence during the $t_{\mathrm{b}}$ evolution periods must be increased in duration by $90^{\circ}$. As a result of this phase change, the effective precession axis is altered, and so the compensation pulse at the end of the evolution pulse must also be altered so that the $45^{\circ}$ pulse is applied along the $x$ axis. Figure 3 shows the effective precession axes for the two MREV8 sequences together with the corresponding compensation pulse that needs to be applied.

Using Eqs. [19] and [20], various aspects of the MAH experiment were simulated to see if the approach taken for the phase cycling would work. To begin with, a simple MAH experiment was simulated without any homonuclear multiplepulse dipolar decoupling. From this basic program, two further programs were written which incorporate a homonuclear dipolar decoupling MREV8 sequence into the (a) evolution and (b) acquisition periods of the sequence. By separating this step into two programs, the effectiveness of the proposed phase cycling schemes on the evolution and acquisition periods is investi-



| Preparation Pulse <br> Phase | MREV8 Sequence <br> Phases | Normalized <br> Precession Axis | $45^{\circ}$ Quadrature <br> Compensation Pulse |
| :---: | :---: | :---: | :---: |
| $X$ | $(X \bar{Y})(\bar{Y} \bar{X})(\overline{X Y})(Y X)$ | $\frac{1}{\sqrt{2}}(1,0,1)$ | $Y$ |
| $Y$ | $(X X)(\overline{X Y})(\bar{Y} X)(\bar{X} Y)$ | $\frac{1}{\sqrt{2}}(0.1 .1)$ | $X$ |

FIG. 3. Effective precession axes under MREV8 multiple-pulse decoupling as a function of the preparation pulse phase. Listed below are MREV8 sequence phases, the normalized precession axis, and the $45^{\circ}$ compensation pulse $c$ required for quadrature preservation during incorporation into the MAT sequence.


FIG. 4. ${ }^{19} \mathrm{~F}$ MP-MAT NMR spectrum ( 188.2976 MHz ) of AgTFA (a) collected at $\nu_{\text {rot }}=46 \mathrm{~Hz}\left(\tau_{\text {rot }}=21.739 \mathrm{~ms}\right)$ using the pulse sequence of Fig. 1: 20 time increment $\Delta t_{\mathrm{b}}, 32$ FIDs for each increment with a recycle delay of 6 s . The evolutionary MREV8 cycle time $=12 \times 3.0 \mu \mathrm{~s}=36.0 \mu \mathrm{~s}$; each $\Delta t_{\mathrm{b}} / 3$ evolution increment was composed of two MREV8 sequences for a total dwell time of $2 \times 3 \times 36.0 \mu \mathrm{~s}=216.0 \mu \mathrm{~s}$. The total evolution time was $20 \times 216.0$ $\mu \mathrm{s}=4320 \mu \mathrm{~s}$. The acquisition MREV8 cycle time $=\Delta t_{\mathrm{a}}=12 \times 3.4 \mu \mathrm{~s}=$ $40.8 \mu \mathrm{~s}$, and 256 acquisition points were collected. Total experimental time was 2.1 h . Pulses are as in Fig. 2. (b) A simulation using the MAH model as described in text. For both plots, the floor of the contour plot was set to $10 \%$ of $I_{\max }$ with 9 contours at $10 \% I_{\max }$ intensity increments. Sum projections are given in both dimensions, and the chemical shift scale is corrected for MREV8 scaling of 0.499 .
gated without interference from each other. From these two simulations it was clear that the phase cycling did allow the incorporation of the MREV8 sequence into the MAH experiment. A final program combines the two earlier programs, with the MREV8 sequence in both the acquisition and evolution periods of the MAH experiment. Figure 4b shows a contour plot of a computer simulation of the final MAH experiment for the two-spin case with an MREV8 homonuclear dipolar decoupling sequence in both periods. The isotropic shifts were 1 and -2 kHz , the CSA values were 14.25 and -14.25 kHz (with both asymmetries set to 0.0 ), and the transmitter was set
at 0 Hz . The MREV8 cycle time was $36 \mu \mathrm{~s}$ in both the $t_{\mathrm{a}}$ acquisition and the $t_{\mathrm{b}}$ evolution dimensions. The step increment in the evolution dimension was set equal to a single MREV8 cycle, and $64 t_{\mathrm{b}}$ evolution increments were calculated. Relaxation of in-plane magnetization during the storage period is simulated by the setting of the off-diagonal elements of the density matrix to zero. $T_{2}$ relaxation during the acquisition and evolution periods is performed by the application of a weighting function when the data were processed. Ideal delta pulses were used in the simulation. The figure indicates that the experiment retains quadrature in both dimensions and that a pure adsorption spectrum is obtained. The small glitch at zero frequency in the anisotropic dimension arises from the variability of the precession axis of the magnetization brought about by the large chemical shift offsets (38).

## (c) The Multiple-Pulse Magic-Angle-Turning Experiment

The ${ }^{19}$ F MP-MAT NMR spectrum of AgTFA. The focus of the initial MP-MAT experiments was to examine the feasibility of the technique using AgTFA. AgTFA has a single peak in the 1-dimensional ${ }^{19}$ F MAS NMR spectrum at $\delta=-71.5 \mathrm{ppm}$ and a narrow, nearly axially symmetric CSA powder pattern (40). Figure 4 a shows the ${ }^{19} \mathrm{~F}$ MP-MAT spectrum of AgTFA with sum projections along the $F_{2}$ acquisition (horizontal) and $F_{1}$ evolution (vertical) dimensions. The projections of both dimensions demonstrate the removal of the ${ }^{19} \mathrm{~F}-{ }^{19} \mathrm{~F}$ homonuclear dipolar interaction, giving 2-D spectral features arising from the chemical shift interaction alone. The pseudo-static lineshape shows a nearly axially symmetric CSA powder pattern, with values of the principal components of the chemical shift $\left(\delta_{11}=+45.3 \mathrm{ppm}, \delta_{22}=-16.7 \mathrm{ppm}, \delta_{33}=-28.7 \mathrm{ppm}\right.$ with respect to $\delta_{\text {iso }}$ ) similar to those in the literature $\left(\delta_{11}=+49.4\right.$ $\left.\mathrm{ppm}, \delta_{22}=\delta_{33}=-24.7 \mathrm{ppm}\right)(40)$. The slight asymmetry of the current result may be a result of the higher resolution afforded by the larger magnetic field (4.7 T vs 1.35 T$)$. The deviation from axial symmetry for a rapid rotating $-\mathrm{CF}_{3}$ group may be related to molecular packing effects.

The sum projection along the $F_{1}$ isotropic evolution dimension shows that the MP-MAT experiment has removed both the homonuclear dipolar interaction and the chemical shift anisotropy. The single peak at $\delta=-71.5 \mathrm{ppm}$ has a full width at half height (FWHH) of 4.0 ppm , which is greatly reduced from the sum projection along the $F_{2}$ anisotropic acquisition dimension ( $\xi=68 \mathrm{ppm}$ ). The MREV8 multiple-pulse decoupling sequence averages the homonuclear dipolar interaction to zero and partially averages the chemical shift interaction. The chemical shift, then, is scaled under MREV8 conditions by a theoretical value of $\sqrt{2} / 3=0.471$; the actual value depends on factors such as the pulse duration, pulse imperfections, and MREV8 cycle times. The scaling factor can be determined experimentally by measuring the position of the isotropic peak as a function of the transmitter offset. Four MP-MAT experiments were carried out with the transmitter frequency set to values $-1,1,2$, and 3 kHz from the ${ }^{19} \mathrm{~F}$ resonance frequency.


FIG. 5. (a) Determination of MREV8 scaling factor for the evolution dimension of the ${ }^{19} \mathrm{~F}$ MP-MAT spectra of AgTFA. The ${ }^{19} \mathrm{~F}$ transmitter was positioned $-1,1,2$, and 3 kHz from resonance, and the corresponding evolution dimension frequency offsets were measured. The slope of the fit gives a scaling factor of $0.499 \pm 0.002$. (b) FWHH linewidths of the ${ }^{19} \mathrm{~F}$ isotropic chemical shift sum projection as a function of sample rotation frequency collected using the MP-MAT pulse sequence (O); MP-MAT ${ }^{ \pm}$, no rotational echo extensions in $t_{\mathrm{b}}(\triangle)$; MP-MAT ${ }^{ \pm}$, one rotational echo extension in $t_{\mathrm{b}}(\square)$; MP-MAT ${ }^{ \pm}$, two rotational echo extensions in $t_{\mathrm{b}}(\diamond)$; MP-MAT ${ }^{ \pm}$, three rotational echo extensions in $t_{\mathrm{b}}(\nabla)$. The MP-MAT ${ }^{ \pm}$spectra were collected at $408 \mathrm{~Hz}\left(\tau_{\text {rot }}=2.451 \mathrm{~ms}\right)$ using the pulse sequence depicted in Fig. 7: $188.29225 \mathrm{MHz}, 40$ time increments $\Delta t_{\mathrm{b}}, 48$ transients for each increment with a 6-s recycle delay. The evolutionary MREV8 cycle time $=12 \times 3.4$ $\mu \mathrm{s}=40.8 \mu \mathrm{~s}$; each $\Delta t_{\mathrm{b}} / 3$ evolution increment was composed of one MREV8 sequence for a total dwell time of $3 \times 40.8 \mu \mathrm{~s}=122.4 \mu \mathrm{~s}$ and a total evolution time of $4896 \mu \mathrm{~s}$. The acquisition MREV8 cycle time $=\Delta t_{\mathrm{a}}=12 \times 3.4 \mu \mathrm{~s}=$ $40.8 \mu \mathrm{~s}$, and 272 acquisition points were collected. Pulses: $45^{\circ}=0.80 \mu \mathrm{~s} ; 90^{\circ}$ $=1.36 \mu \mathrm{~s} ; 135^{\circ}=1.87 \mu \mathrm{~s}$. In all cases, the chemical shift scale is corrected for MREV8 scaling of 0.499 .

The position of the isotropic peak in the evolution dimension was measured with respect to the transmitter position, and the data are plotted in Fig. 5a. The slope of the linear fit gives the scaling factor as $0.499 \pm 0.002$. The values for the FWHH of the isotropic peak were invariant to the transmitter offset.

The effective evolution spectral width, already reduced by a factor of 2 by the MREV8 scaling factor, is reduced by a further factor of 3 by the MAT component of the experiment according to Eq. 1. This restricted spectral width in the evolution dimension, in principle, limits the extent to which CSA patterns of vastly different resonance frequencies can be separated. Of the three naturally abundant spin $-\frac{1}{2}$ nuclei, the ${ }^{19} \mathrm{~F}$ nucleus, with its large chemical shift interaction, is most sensitive to this limitation. Nevertheless, with evolutionary MREV8 cycles times as short as $24 \mu s$, an effective evolutionary spectral width of 41.7 kHz is achievable which, at a field strength of 4.7 T , means a chemical shift range of $\sim 75 \mathrm{ppm}$ after accounting for the MREV8 scaling factor.

The ${ }^{19}$ F MP-MAT NMR spectrum of a mixture of AgTFA and calcium difluoride. A physical mixture of AgTFA ( 0.442 g , $3.82 \mathrm{mmol})$ and $\mathrm{CaF}_{2}\left(0.234 \mathrm{~g}, 3.00 \mathrm{mmol}, \delta_{\text {iso }}=-106 \mathrm{ppm}\right)$ was prepared to give a model two-site system to demonstrate
that the MP-MAT technique is applicable to multiple-site cases. The isotropic chemical shifts of the two components differ by 35 ppm (41). Since there is no anisotropy in the $\mathrm{CaF}_{2}$ chemical shift, the MP-MAT spectrum should indicate whether there is any intensity contamination between different isotropic signals.

Figure 6 shows the ${ }^{19} \mathrm{~F}$ MP-MAT NMR contour plot of the AgTFA/calcium difluoride mixture with sum projections in both dimensions. Two peaks in the $F_{1}$ isotropic evolution dimension are clearly resolved, with FWHH values of 680 (AgTFA) and $530\left(\mathrm{CaF}_{2}\right) \mathrm{Hz}$. The transmitter has been placed between the two peaks at 188.294273 MHz . The absence of ghost or pedestal artifacts in both dimensions, often seen with multiple-pulse sequences, demonstrates that the addition of magnetization developed under MREV8 conditions is consistent with the MAT requirements.

## (d) Resolution Enhancement of the Isotropic Dimension

For the separation of CSA powder patterns of very similar isotropic chemical shifts, the resolution in the $F_{1}$ evolution dimension is crucial. The value for the AgTFA linewidth of the $F_{1}$ sum projection in the ${ }^{19} \mathrm{~F}$ MP-MAT experiment depicted in Fig. 4 is 450 Hz . This FWHH is much larger than similar measurements with AgTFA under 1-dimensional MAS conditions (FWHH $=21 \mathrm{~Hz}$ at $\left.\nu_{\text {rot }}=2249 \mathrm{~Hz}\right)$. Attempts to narrow the linewidth of a stationary sample of powdered $\mathrm{CaF}_{2}$ by adjustments of the magnet shims were unsuccessful. However, when the $\mathrm{CaF}_{2}{ }^{19} \mathrm{~F}$ spectrum was collected under MAS conditions, the linewidth dropped considerably in value. The above observations indicate that the broadening of the ${ }^{19} \mathrm{~F}$ peak arises


FIG. 6. ${ }^{19}$ F MP-MAT NMR spectrum ( 188.2976 MHz ) of AgTFA/calcium difluoride mixture collected at a rotor spinning speed of $71.5 \mathrm{~Hz}\left(\tau_{\text {rot }}=\right.$ 13.986 ms ). Fifty $\Delta t_{\mathrm{b}} / 3$ increments of a $36.0-\mu \mathrm{s}$ MREV8 cycle were collected (total dwell time $=108.0 \mu \mathrm{~s}$ ) for a total evolution time of $5400 \mu \mathrm{~s}$. Each $t_{\mathrm{a}}$ acquisition was collected 16 times using a $40.8 \mu$ s MREV8 cycle with 256 points and using a recycle delay of 6 s ; the total experimental time was 2.7 h . The floor and ceiling were set to 5 and $85 \% I_{\max }$, respectively, with contour increments of $10 \% I_{\max }$. The $F_{1}$ and $F_{2}$ spectra are the sum projections. The chemical shift scale is corrected for MREV8 scaling using a value of 0.499.


FIG. 7. The MP-MAT ${ }^{ \pm}$pulse sequence resulting from the incorporation of MREV8 multiple pulse homonuclear decoupling into the MAT ${ }^{ \pm}$sequence of Gan and Ernst (6). Note that the "plus" and "minus" variants differ in the acquisition of the $t_{\mathrm{a}}$ FID as well as in the phases of the receiver and pulses $p_{x}$. The phase table is given in Table 3. MREV8 multiple-pulse decoupling in the two dimensions are as in Figs. 1b and 1c.
from the inhomogeneity of the magnetic field at the sample due to bulk magnetic susceptibility effects. We now report several strategies to reduce the linewidth in the evolution dimension and thus increase the resolution of the 2-D MP-MAT experiment.

Sample spinning. ${ }^{19} \mathrm{~F}$ MP-MAT NMR experiments were carried out using AgTFA at sample spinning speeds between $\nu_{\text {rot }}=45$ and 408 Hz . The values of the FWHH for the summed $F_{1}$ projections as a function of the frequency of sample magicangle rotation are reported in Fig. 5b as open circles. The linewidths drop considerably to 210 Hz at $\nu_{\text {rot }}=408 \mathrm{~Hz}$. Due to the restriction of the total evolution time to one rotor period, faster spinning speeds will lead to truncation of the evolution signal. Furthermore, at MAS speeds greater than the ${ }^{19} \mathrm{~F}$ natural linewidth, a spinning-sideband manifold rather than a pseudostatic CSA pattern is collected.

The MP-MAT ${ }^{ \pm}$pulse sequence. Additional resolution enhancement is achieved by modification of the MP-MAT pulse sequence according to the MAT ${ }^{ \pm}$pulse sequence of Gan and Ernst (6). Rather than collecting three $t_{\mathrm{b}}$ evolutionary increments, two are collected, and the sequence generates a sheared spectrum. A shearing transformation is easily applied in subsequent data processing to give a spectrum with the CSA patterns parallel to the acquisition dimension. Furthermore, if the $t_{\mathrm{a}}$ and $t_{\mathrm{b}}$ increments are synchronized to an integral multiple of the rotor period, rotational echoes in the acquisition dimension can be used to extend the time-domain signal of the evolution dimension. The consequence of this data shifting is enhanced digital resolution in the evolution dimension. The
${ }^{19} \mathrm{~F}$ MAT EXPERIMENTS WITH MULTIPLE-PULSE DECOUPLING
tAbLE 3
The Basic Phase Table for the MP-MAT ${ }^{ \pm}$Pulse Sequence in Fig. 7

| $n t$ | $F_{1}$ |  |  |  | $F_{2}$ |  |  |  |  |  |  | $F_{3}$ |  |  |  |  |  |  | $F_{4}$ |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | 1 | 2 | 3 | 4 | 5 |  | 6 |  | 7 |  | 8 |  | 9 |  | 10 |  | 11 | 12 | 13 | 14 | 15 | 16 |
| $r$ | $Y$ | $Y$ | $Y$ | $Y$ | $Y$ |  | $Y$ |  | Y |  | $Y$ |  | $Y$ |  | $Y$ |  | $Y$ | $Y$ | $Y$ | Y | Y | Y |
| $p_{1}$ | $\bar{Y}$ | $\bar{Y}$ | $\bar{Y}$ | $\bar{Y}$ | $X$ |  | $X$ |  | $X$ |  | $X$ |  | $\bar{Y}$ |  | $\bar{Y}$ |  | $\bar{Y}$ | $\bar{Y}$ | $X$ | $X$ | $X$ | $X$ |
| $p_{2}$ | $\bar{Y}$ | $\bar{Y}$ | $\bar{Y}$ | $\bar{Y}$ | X |  | $X$ |  | $X$ |  | $X$ |  | $X$ |  | $X$ |  | $X$ | X | $\bar{Y}$ | $\bar{Y}$ | $\bar{Y}$ | $\bar{Y}$ |
| $r_{\phi}$ | $\{135$ | 45 | 90 | 90 | 135 |  | 45 |  | 90 |  | 90 |  | 135 |  | 45 |  | 90 | 90 | 135 | 45 | 90 | 90 |
|  | \{ Y | $\bar{Y}$ | $X$ | $\bar{X}$ | Y |  | $\bar{Y}$ |  | $X$ |  | $\bar{X}$ |  | Y |  | $\bar{Y}$ |  | $X$ | $\bar{X}$ | Y | $\bar{Y}$ | $X$ | $\bar{X}$ |
| $r c \nu r$ | $Y$ | $\bar{Y}$ | $X$ | $\bar{X}$ | $Y$ |  | $\bar{Y}$ |  | $X$ |  | $\bar{X}$ |  | $Y$ |  | $\bar{Y}$ |  | $X$ | $\bar{X}$ | $Y$ | $\bar{Y}$ | $X$ | $\bar{X}$ |
| Sum | $X$ |  |  |  | $\bar{X}$ |  |  |  |  |  |  |  | $\bar{Y}$ |  |  |  |  |  | $\bar{Y}$ |  |  |  |
|  | $F_{5}$ |  |  |  | $F_{6}$ |  |  |  |  |  |  | $F_{7}$ |  |  |  |  |  |  | $F_{8}$ |  |  |  |
| $n t$ | 1 | 2 | 3 | 4 | 5 | 6 |  | 7 |  | 8 |  | 9 |  | 10 |  | 11 |  | 12 | 13 | 14 | 15 | 16 |


| Sum | $X$ | $\bar{x}$ | $Y$ |
| :--- | :---: | :---: | :---: |

Note. The $P^{+}$(top) and $P^{-}$(bottom) data sets are collected with variation of the required phase for the sum $P^{+}=\sum_{i=1}^{4} F_{i}$ and $P^{-}=\sum_{i=5}^{8} F_{i}$; nt represents the transition number.

MP-MAT ${ }^{ \pm}$sequence is depicted in Fig. 7 and the pulse phases are recorded in Table 3. Each of the eight different FIDs were collected independently using the same receiver phase and combined to give the real and imaginary FIDs as described above.

The improvement in the AgTFA linewidths at $\nu_{\text {rot }}=408 \mathrm{~Hz}$ is dramatic, as indicated by the noncircle symbols in Fig. 5b. The MP-MAT ${ }^{ \pm}$sequence gives an immediate improvement relative to the MP-MAT sequence at $\nu_{\text {rot }}=408 \mathrm{~Hz}$ which may arise from the development of the evolution signal over two rather than three evolutionary periods. As the length of the evolution signal is extended from 4896 to $19584 \mu$ s by rotational echo data shifting, the linewidth drops to 50 Hz .

Resolution enhancement using the rotational echo $t_{\mathrm{b}}$ extension technique also increases the noise in the ${ }^{19} \mathrm{~F} \mathrm{MP}^{2} \mathrm{MAT}^{ \pm}$as indicated in the sum $F_{1}$ projections in Fig. 8. Ridge features, reminiscent of spinning sidebands, lie parallel to the $F_{2}$ axis and increase in prominence as the number of rotational echo extensions of the evolutionary signal increase. These features have been described by Grant and co-workers for the 5- $\pi$ MAT and FIREMAT sequences which employ evolutionary block replication techniques (11,12). In the present case, ridge features arising from the difference in the decay of magnetization intensity along the $t_{\mathrm{a}}$ acquisition and $t_{\mathrm{b}}$ evolution dimensions are understood in terms of a sinex/x effect resulting from truncation of the evolution time signal in discrete blocks. In the acquisition dimension, magnetization decays as $\exp \left(-t_{\mathrm{a}} / T_{2}\right)$, and as $\exp \left(-t_{\mathrm{b}} / T_{2}\right)$ in the evolution dimension, $T_{2}$ being the transverse magnetization relaxation time constant. The extended evolution signal is created by taking the first rotational echo, which has an intensity of $I_{\mathrm{a}}=I_{0} \exp \left(-\tau / T_{2}\right)$, and adding it to the evolution dimension terminus at time $2 \tau / 3$, which has an intensity $I_{\mathrm{b}}=I_{0} \exp \left(-(2 \tau / 3) / T_{2}\right)$. Clearly, $I_{\mathrm{b}}>$ $I_{\mathrm{a}}$; similar intensity differences occur at each rotational echo
extension at times $2 k \tau / 3$, where $k=$ the number of rotational echoes. The resulting extended $t_{\mathrm{b}}$ evolutionary signal is characterized by a stepped decay function which, in turn, leads to the observed sinex/x artifacts.

In theory, the decay of magnetization intensity in the evolution dimension will step down as a function of $k$ rotational echo extensions. Furthermore, the noise of the evolution signal in each block may add coherently as a function of $k$. This feature becomes more pronounced when $T_{2}$ is of the order of or less than the rotor period $\tau_{\text {rot }}$. In practice, however, errors in rotor synchronization and MREV8 pulse imperfections contribute to additional loss of signal magnetization and cause the intensity of the extended evolution signal to step up as a function of $k$. Both truncation and noise-coherence effects can be treated in part by application of an exponential damping function to the acquisition domain before rotational echo extension of the $t_{\mathrm{b}}$ signal. While this treatment reduces the prominence of the artifact signals, it also decreases the digital resolution and hence increases the linewidth of the resulting signals (Fig. 8d).

Evolution MREV8 cycle time $t_{c}$. In 1-dimensional multi-ple-pulse NMR experiments, the cycle time $t_{\mathrm{c}}$ is determined by the $90^{\circ}$ pulse duration and the $\tau$ delay between pulses. To achieve a large spectral width, $t_{\mathrm{c}}$ should be as short as possible. Since the signal must be sampled at least once during the cycle, the probe ring-down and dead-time characteristics determine the minimum multiple-pulse $\tau$ value and hence $t_{c}$. Short $t_{\mathrm{c}}$ values are also desirable to improve the effectiveness of the homonuclear dipolar averaging.

Indirect sampling of the evolutionary signal allows multiplepulse cycle times to be shorter than in the directly sampled acquisition dimension. In addition to increasing the spectral width, use of a $\tau$ delay of $2.0 \mu \mathrm{~s}\left(t_{\mathrm{c}}=24.0 \mu \mathrm{~s}\right)$ reduces slightly


FIG. 8. Resolution enhancement in the sum isotropic chemical shift projections of the ${ }^{19} \mathrm{~F}$ 2-D isotropic chemical shift versus CSA spectrum of AgTFA. (a) MP-MAT, $\nu_{\text {rot }}=71.5 \mathrm{~Hz}$, (b) MP-MAT ${ }^{ \pm}, \nu_{\text {rot }}=408 \mathrm{~Hz}$ with no rotational echo extensions in $t_{\mathrm{b}}$, (c) MP-MAT ${ }^{ \pm}, \nu_{\text {rot }}=408 \mathrm{~Hz}$ with two rotational echo extensions in $t_{\mathrm{b}}$, (d) MP-MAT ${ }^{ \pm}$, $\nu_{\text {rot }}=408 \mathrm{~Hz}$ with 200 Hz exponential linebroadening applied to the acquisition dimension prior to two rotational echo extensions in $t_{\mathrm{b}}$.
the linewidth in the $t_{\mathrm{b}}$ dimension of AgTFA at $\nu_{\mathrm{rot}}=45 \mathrm{~Hz}$ to 330 Hz compared to 350 Hz using a $\tau$ delay of $3.0 \mu \mathrm{~s}\left(t_{\mathrm{c}}=\right.$ $36.0 \mu \mathrm{~s})$. It should be feasible to incorporate more sophisticated multiple-pulse decoupling schemes such as BR24 (42) and CORY24 (36) as well as windowless and semi-windowless variants. It is anticipated that these modifications will contribute to an improvement in resolution.

Final comments on resolution enhancement. Three approaches to improve the resolution of the MP-MAT experiment have been investigated: (a) increased sample rotation, (b) improvements in the MAT component of the experiment, and (c) shorter multiple-pulse $t_{\mathrm{c}}$ cycle times. Each strategy has led to resolution enhancement as detailed above. It is noted, however, that these three investigations by no means exhaust the list of potential improvements to the technique. Grant and co-workers have shown the benefit of a rotor synchronization strategy by which the MAT pulses are triggered by optical sensing of the
sample rotor position (5). We note that the sample rotation conditions described herein ( $\pm 0.5 \mathrm{~Hz}$ ) give an error in rotor period timing of $\pm 100 \mu \mathrm{~s}$ at a spinning speed of $\nu_{\text {rot }}=71.5 \mathrm{~Hz}$. The error is comparable to the evolutionary dwell time and is likely to give rise to error in the isotropic chemical shift dimension; this error is manifested in (a) a low signal-to-noise ratio and (b) additional magnetization decay resulting in poorly resolved signals.

The present MAT experiments fall into the "projection" class of MAT experiments (11). Isotropic chemical shift and CSA correlation spectra are also obtained from the " $5-\pi$ " class of MAT experiments. The recent FIREMAT version of a $5-\pi$ MAT experiment for dilute spin- $\frac{1}{2}$ nuclei offers a significant improvement in the signal-to-noise ratio (11). Analogous 5- $\pi$ MAT experiments incorporating multiple-pulse decoupling sequences will, in principle, offer similar advantages to abundant spin- $\frac{1}{2}$ nuclei. A $5-\pi$ MP-MAT sequence is more appropriate for nuclei with large CSA values such as ${ }^{19} \mathrm{~F}$ and ${ }^{31} \mathrm{P}$ than for ${ }^{1} \mathrm{H}$, which is better suited to the projection MAT experiment because of the small ${ }^{1} \mathrm{H}$ CSA values.

More efficient homonuclear dipolar decoupling gives narrower linewidths in the $t_{\mathrm{b}}$ isotropic dimension. This improvement may be achieved by decreasing the multiple-pulse cycle time and by employing multiple-pulse dipolar decoupling schemes which average to zero the higher order homonuclear dipolar interaction terms. Such sequences include the BR24 and CORY24 sequences as well as the windowless and semiwindowless versions. One must select compensation pulse durations and phases appropriate to the effective magnetization precession axis.

## CONCLUSIONS

We have demonstrated the use of the MAT experiment on abundant spin- $\frac{1}{2}$ nuclei where the homonuclear dipolar interaction is dominant. This interaction is suppressed though the incorporation of an MREV8 homonuclear dipolar decoupling sequence into the MAT experiment. The quadrature nature of the MAT experiment is preserved, thereby maximizing the spectral width of the experiment and the effectiveness of the MREV8 sequence in suppressing the homonuclear dipolar interaction.

## ACKNOWLEDGMENTS

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[^1]:    Note. nt represents the transition number.

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